

An International journal in English published online quarterly with aim to publish peer reviewed review and research articles in rapidly developing field of pharma and life sciences

**We publish Research / Review article in the following subjects**

**Life Sciences**

**Branches :** Agricultural Science, Biochemistry, Biology, Bioinformatics, Botany, Cytology, Cell biology, Chemistry, Ecology, Endocrinology, Entomology, Environmental Sciences, Food science and Technology, Genetics, Genomics & Proteomics, Immunobiology, Molecular biology, Marine Science, Microbiology, Neurobiology, Pathology, Physics, Physiology, Psychology, Veterinary Science, Zoology

**Pharmaceutical Sciences**

**Branches :** Biotechnology, Clinical and Hospital pharmacy, Herbal technology, Industrial Pharmacy, Immunology, International Regulatory Affairs, Medicine, Neuroscience, Novel drug delivery system, Nanotechnology, Pharmaceuticals, Pharmacology & Toxicology, Pharmacognosy & Phytochemistry, Pharmacy practice, Pharmaceutical Engineering, Pharmaceutical Management, Pharmaceutical Analysis, Pharmaceutical Chemistry.



**Impact Factor \*0.672**



Minneapolis , United States of America



U.S.A



Oxford, United Kingdom



**Friedrich-Alexander-Universität Erlangen-Nürnberg**



Erlangen, Germany



**University of Reading**

United Kingdom



**ULRICHSWEB™**  
GLOBAL SERIALS DIRECTORY  
USA (Washington DC)



Spring Hill College  
Mamie and John Burke memorial library  
USA (Alabama)



USA (Alabama)



USA (connecticut)



United kingdom



Japan



Japan



USA (Massachusetts)



DOSHISHA UNIVERSITY

Japan



Naropa University  
Allen Ginsberg Library  
USA (Colorado)



New York, NY, United States



Jacksonville State University  
Houston Cole Library  
USA (Alabama)



Illinois state university  
Milner Library  
USA (ILLINOIS)



**Huntington College**

Huntington College  
Houghton Memorial library  
USA (Alabama)



IJLPR would take care in making your article published without delay with your cooperation. IJLPR hopes that Researchers, Research scholars, Academician, Industrialists, Consultancy etc. would make use of this journal publication for the development of science and technology.

\*Kindly visit Instruction to authors available at [www.ijlpr.com](http://www.ijlpr.com) for submission of manuscript for publication.

Any feed back / query kindly email to

[editorofijlpr@rediffmail.com](mailto:editorofijlpr@rediffmail.com)

[editorijlpr@yahoo.com](mailto:editorijlpr@yahoo.com)

or you can call +91 9908947749 / **+91 9676175127 / 9676175127**



## RESCUE OF INFLAMMATORY RENAL DAMAGE BY MEDICINAL PLANT EXTRACTS IN DIABETIC RATS

AMNA A SADDIQ<sup>1,2</sup> AND AZZA. M. MOHAMED<sup>\*3,4</sup>

<sup>1</sup>Department of Biology, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia

<sup>2</sup>Department of Biology, Faculty of Science, University of Jeddah, Jeddah, Saudi Arabia

<sup>3</sup>Applied Biochemistry Department, Faculty of Science, University of Jeddah, Jeddah, Saudi Arabia

<sup>4</sup>Therapeutic Chemistry Department, Drug Industry Division, National Research Center, Cairo, Egypt.

### ABSTRACT

Diabetic nephropathy (DN) is the most critical diabetic complication that leads to renal failure. The purpose of this research was to explore the anti-diabetic effectiveness of *Cucumis melo* var. *flexuosus* and/or *Phoenix dactylifera* fruit aqueous extracts and their mechanisms in alleviating nephropathy in hyperglycemic rats. Diabetes was promoted in rats by injection (i.p.) of a single dose of streptozotocin (STZ). *C. flexuosus* and *P. dactylifera* extracts (CFE and PDE respectively) were ingested to diabetic rats for thirty consecutive days. The data revealed that intake of either plant extract or their combination to diabetic rats, significantly diminished the serum glucose and raised the serum insulin concentration. The plant extracts significantly ameliorated the increases in relative kidney weights as well as in renal tumor necrosis factor (TNF- $\alpha$ ), serum C-reactive protein (CRP) and renal vascular endothelial factor (VEGF). Serum kidney function markers (creatinine, uric acid and urea) were also decreased in diabetic rats treated with the plant extracts. Histopathological observation was also carried out to confirm the biochemical results. This study has proven that the current plant extracts have potential hypoglycemic effect and could attenuate diabetic inflammatory renal damage in rats. The combination of the two extracts was synergistically the effective one. This result may help in exploring a novel therapy to manage diabetes and its complications.

**KEY WORDS:** *Phoenix dactylifera*, *Cucumis melo* var. *flexuosus*, inflammatory molecules, renal damage.



AZZA. M. MOHAMED\*

Applied Biochemistry Department, Faculty of Science, University of Jeddah, Jeddah, Saudi Arabia  
Therapeutic Chemistry Department, Drug Industry Division, National Research Center, Cairo, Egypt.

Received on: 04-10-2018

Revised and Accepted on: 03-12-2018

DOI: <http://dx.doi.org/10.22376/ijpbs/lpr.2019.9.1.L24-33>

## INTRODUCTION

Diabetes mellitus ranks among the top causes of death worldwide. This syndrome is identified by impaired of glucose regulation, resulting from deficiency in insulin release, insulin function or both, causing defect in glucose degradation and other energy-producing elements, including fats and proteins<sup>1</sup>. Modification in metabolism leads to microvascular problems, including nephropathy<sup>2</sup>. Diabetic nephropathy (DN) is the principle reason of progressive kidney damage, leading to dialysis or transplantation<sup>3</sup>. DN has been shown to develop in 20–50% of diabetic subjects and has become a leading reason of renal failing globally<sup>3</sup>. DN is discriminated by structural and functional aberrations<sup>4</sup>. Patients with DN have a gradual drop in glomerular function<sup>5</sup>. Studies on DN, revealed that chronic inflammatory stress participates in its development and progression<sup>6</sup>. NF-kB related inflammatory reactions has been shown to have a fundamental role in the pathogenesis of DN<sup>7</sup>. Generation of inflammatory molecules, like growth factors, cytokines and chemokines has been proven in DM<sup>4</sup>. Plant based therapy is presently used for controlling diabetes due to its capability to suppress the incidence and/or progression of its complexity. Cucurbitaceus is a family for many medicinal plants cultivated in equatorial regions<sup>8</sup>. Some reports have demonstrated that many cucurbits have potential pharmacological properties versus complexity of diabetic<sup>9-10</sup>. *Cucumis melo* var. *flexuosus* (known as snake melon) is one of cucurbit plants<sup>11</sup>. *C. melo* var. *flexuosus* leaf extract has been found to possess antidiabetic, anti-inflammatory, antioxidant and anti-apoptosis capacities<sup>12</sup>. *Phoenix dactylifera* Linn. (date palm, family *Arecaceae*) fruits are broadly consumed worldwide for their beneficial nutritive importance. The fruit water extract has many medicinal activities, such as anti-mutagenic, anti-inflammatory, antioxidant and antidiabetic activities<sup>13-14</sup>. The anti-inflammatory effect attributes to its active components namely flavonoids, steroids, saponins and phenolic compounds<sup>15</sup>. It has reported that date can stimulate insulin release and suppress glucose absorption<sup>16</sup>. Although the antidiabetic impacts of both *C. melo* var. *flexuosus* and *P. dactylifera* were studied, the exact mode of actions of their extracts in management diabetic renal damage are still unexplored. The goal of this work was to investigate the key mechanism (s) of CFE and /or PDE in protecting against diabetes promoted renal damage in diabetic rats.

## MATERIALS AND METHODS

### Chemicals

All chemicals utilized in this study were highly pure, obtained from Sigma and Merck companies, USA.

### Plants

*C. melo* var. *flexuosus* and *P. dactylifera* fruits were bought from the market. The plants were described by a taxonomist in the Department of Biological Science, King Abdulaziz University, Jeddah, Saudi Arabia.

### Procedure of *C. flexuosus* fruit extraction

*C. melo* var. *flexuosus* fruits were cut into pieces after removing the seeds and then dried at 40 °C for 48 hours. 500 g of the dried fruits were crushed and soaked in 4 L bi-distilled water and then heated at 100 °C for half hour. The aqueous phase was gotten by centrifugation for 20 min at 5000xg and then freeze dried.

### Procedure of *P. dactylifera* fruit extraction

*P. dactylifera* L. fruits (400 g) were blended with 4 L bi-distilled water, utilizing an electric blender. The commixture was left for 24 h, and then centrifuged for 25 min. at 4000xg. The liquid phase was gathered and then freeze dried.

### Animals

Male Wistar rats (180-200 g) were used for this investigation. The rats were bought from the Experimental Animal Care Center, King Fahad Medical Research Center, King Abdulaziz University (Jeddah, KSA). Animals were maintained at control circumstances at 22–25 °C and supplied with balanced diet and water *ad libitum*. Animal care was performed following the roles supplied by the Experimental Animal Care Committee (Reference No 1007-16) of Faculty of Science, Jeddah University.

### Experimental design

Diabetes was promoted in diabetic rat groups by injecting a single dose (45 mg/ kg b.w. i. p.) of streptozotocin (STZC, Sigma, USA), prepared in 0.05M citrate buffer (pH 4.5)<sup>17</sup>. The non-diabetic normal group were injected with an equivalent amount of citrate buffer. Blood glucose was monitored utilizing a glucometer, at the tenth day after STZC administration; rats with fasting blood glucose above 220 mg/dl were selected as diabetic. The rats were categorized into five groups (n= 10), group 1, control rats; group 2, diabetic rats; group

3, diabetic rats ingested with CFE (400mg /kg)<sup>9</sup>; group 4, diabetic rats ingested with PDE (400mg/kg)<sup>18</sup>; group 5, diabetic rats ingested with the combination of CFE and PDE (400 mg/ kg each). Lyophilized CFE and PDE were solved in distilled water and given orally to animals for a month, forty days after promotion of diabetes. The combination of the two plant extracts was dissolved as a mixture in distilled water before administration. Each rat was weighed at the beginning and the end of the experiment to estimate the change in body mass. Finally, the rats were fasted for 12-14 hours and the blood samples were withdrawn for serum separation and used for biochemical investigations. After withdrawing the blood, all rats were sacrificed under light anesthesia and the kidney samples were collected and washed in phosphate buffered saline, blotted on a filter paper and weighed then used for biochemical and histopathological tissue analysis.

#### ***Estimation of diabetic and kidney function indices***

Serum fasting glucose level, uric acid, creatinine and urea were estimated using an automatic analyzer (ci16200, Abbott, USA). Insulin was estimated by rat insulin ELISA kit (BioVendor company, Laboratorni medicina a.s. Karasek 1767/1, 621 00 Brno Czech Republic)

#### ***Determination of inflammatory and angiogenic biomarkers***

Tumor necrosis factor (TNF)- $\alpha$ , and vascular endothelial growth factor (VEGF) were determined in kidney tissue, using commercially available ELISA kit in accordance to the guidance provided by the manufacturer (R&D Systems, USA). Serum C-reactive protein (CRP) was measured utilizing latex-enhanced immunonephelometry method (Siemens Dade Behring, Germany).

#### ***Histopathological examination***

Small segments of kidney were treated with 10 % buffered formalin for fixation and then dehydrated and implanted in a paraffin wax. The kidney specimens were cut into sections (3–4- $\mu$ m), stained with hematoxylin-eosin and examined by a light microscope.

## **STATISTICAL ANALYSIS**

The results of the current study were represented as the mean  $\pm$  SD. Statistical analysis was carried out utilizing one-way analysis of variance (ANOVA) followed by Bonferroni as a post-ANOVA test. The

changes among the values were statistically significant at  $p \leq 0.05$ .

## **RESULTS**

#### ***Effect of plant extracts on hyperglycemia markers***

Hyperglycemic rats showed a marked raise in serum glucose and a drop in insulin content versus control animals (Table 1). Ingestion of CFE and /or PDE, markedly ameliorated the glucose and the insulin levels versus diabetic untreated rats ( $P \leq 0.001$ ).

#### ***Effect of plant extracts on kidney hypertrophy***

Diabetic rats had decreased body weights and increased relative kidney weights (kidney hypertrophy) compared with normal rats (Table 2). Administration of CFE and /or PDE appreciably restored the animal body weights and reduced the renal weight in the diabetic treated rat group. Treatment with the combination of CFE and PDE was the effective one in ameliorating the rat body weights and reducing the kidney hypertrophy.

#### ***Effect of plant extracts on inflammatory markers***

As shown in Figures 1 and 2 respectively, the diabetic animals displayed marked increase in the levels of serum CRP and renal TNF- $\alpha$  as compared with the control rats, this deterioration was reversed after the plant extracts administration.

#### ***Effect of plant extracts on renal angiogenesis***

Figure 3 shows a significant increase in the angiogenic factor (VEGF) in the renal of diabetic rats with relation to the control ones ( $P \leq 0.001$ ). Intake of the plant extracts lonely or in a combination, markedly reversed the elevation in this marker.

#### ***Impact of plant extracts on kidney function***

Diabetic rats showed increases in serum levels of kidney function markers (uric acid, creatinine and urea) in comparison with the control rats. CFE and /or PDE treatment, pronouncedly reduced serum kidney function markers (Table 3).

#### ***Effect of plant extracts on histopathological changes***

The impacts of the two plant extracts lonely or in a combination on renal histomorphological changes in the diabetic rats are presented in Figure 4. Diabetic rats showed significant vacuolar degeneration of tubules (Figure 4b); glomerular degeneration (Figure 4c) and infiltration of

inflammatory immune cells (Figure 4d). While, treatment of diabetic rats with CFE and/or PDE (Figures 4e, f & g respectively), restored the normal

structural and morphological architecture of renal glomeruli and tubules with respect to the diabetic rats.

**Table 1**  
*Effect of CFE and /or PDE on hyperglycemic indices in serum of diabetic rat groups*

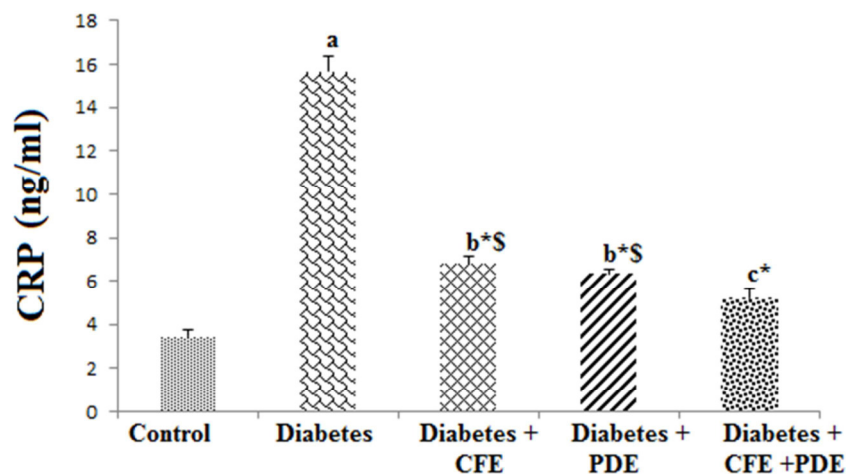
Parameters	Control	Diabetes	Diabetes + CFE	Diabetes + PDE	Diabetes + CFE+ PDE
Glucose (mg/dl)	80.57±3.67	285.77±16.75 <sup>a</sup>	130.94±5.5 <sup>b*</sup>	100.67±7.3 <sup>c*§</sup>	85.9±5.9 <sup>*</sup>
Insulin (pg/ml)	360.36±12.55	125.67±7.27 <sup>a</sup>	245.7±10.5 <sup>a*§</sup>	230.75 ± 6.7 <sup>a§</sup>	289.56± 12.5 <sup>b*</sup>

Data are presented as mean ± S.D. (n=10). Significant difference at: <sup>a</sup>P ≤ 0.001, <sup>b</sup>P ≤ 0.01, <sup>c</sup>P ≤ 0.05 compared with the control group; <sup>\*</sup>P ≤ 0.001 compared with diabetic group; <sup>§</sup>P ≤ 0.05, compared with the combination group (CFE+ PDE).

**Table 2**  
*Effect of CFE and /or PDE on renal hypertrophy in diabetic groups.*

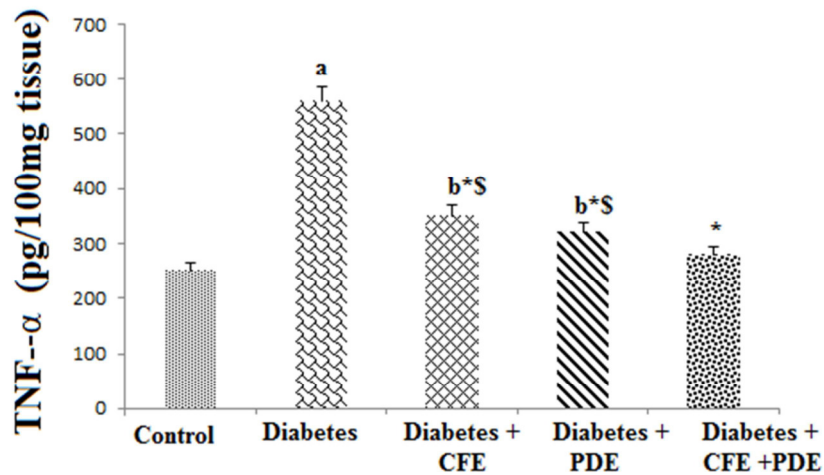
Parameters	Control	Diabetes	Diabetes + CFE	Diabetes + PDE	Diabetes + CFE+ PDE
Body weight change(g)	+70.43±5.17	-10.40±1.2 <sup>a</sup>	+ 50.90±5.30 <sup>c*§</sup>	+ 60.76±3.70 <sup>b*§</sup>	+65.80±4.50 <sup>*</sup>
Relative kidney weight (g /100g body weight)	0.2±0.07	0.49±0.05 <sup>a</sup>	0.29±0.06 <sup>b*§</sup>	0.27±0.07 <sup>b*§</sup>	0.22±0.03 <sup>*</sup>

Data are presented as mean ± S.D. (n=10). Significant difference at: <sup>a</sup>P ≤ 0.001, <sup>b</sup>P ≤ 0.01, <sup>c</sup>P ≤ 0.05; <sup>\*</sup>P ≤ 0.001 compared with diabetic group; <sup>§</sup>P ≤ 0.05, compared with the combination group (CFE+ PDE).



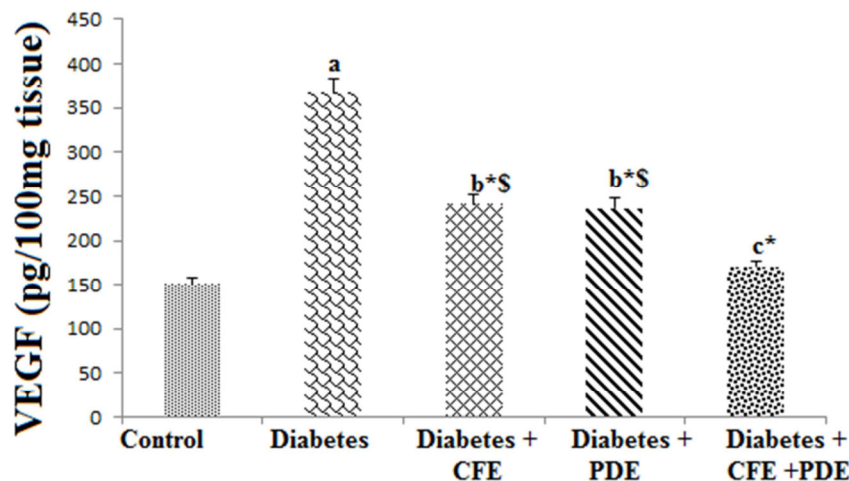
Values are represented as mean±SD (n=10); <sup>a</sup>P ≤ 0.001, <sup>b</sup>P ≤ 0.01, <sup>c</sup>P ≤ 0.05 compared with the control group, <sup>\*</sup>P ≤ 0.001 compared with diabetic group, <sup>§</sup>P ≤ 0.05 compared with combination group (CFE+ PDE).

**Figure 1**  
*Effect of daily oral administration of CFE and /or PDE on the level of serum CRP in diabetic rats.*



Values are represented as mean  $\pm$ SD (n=10); <sup>a</sup>P  $\leq$  0.001, <sup>b</sup>P  $\leq$  0.01 compared with the control group, <sup>\*</sup>P  $\leq$  0.001 compared with diabetic group, <sup>S</sup>P  $\leq$  0.05 compared with combination group (CFE+ PDE).

**Figure 2**  
Effect of daily oral administration of CFE and /or PDE on the level of renal TNF- $\alpha$  in diabetic rats.



Values are represented as mean  $\pm$ SD (n=10); <sup>a</sup>P  $\leq$  0.001, <sup>b</sup>P  $\leq$  0.01, <sup>c</sup>P  $\leq$  0.05 compared with the control group, <sup>\*</sup>P  $\leq$  0.001 compared with diabetic group, <sup>S</sup>P  $\leq$  0.05 compared with combination group (CFE+ PDE).

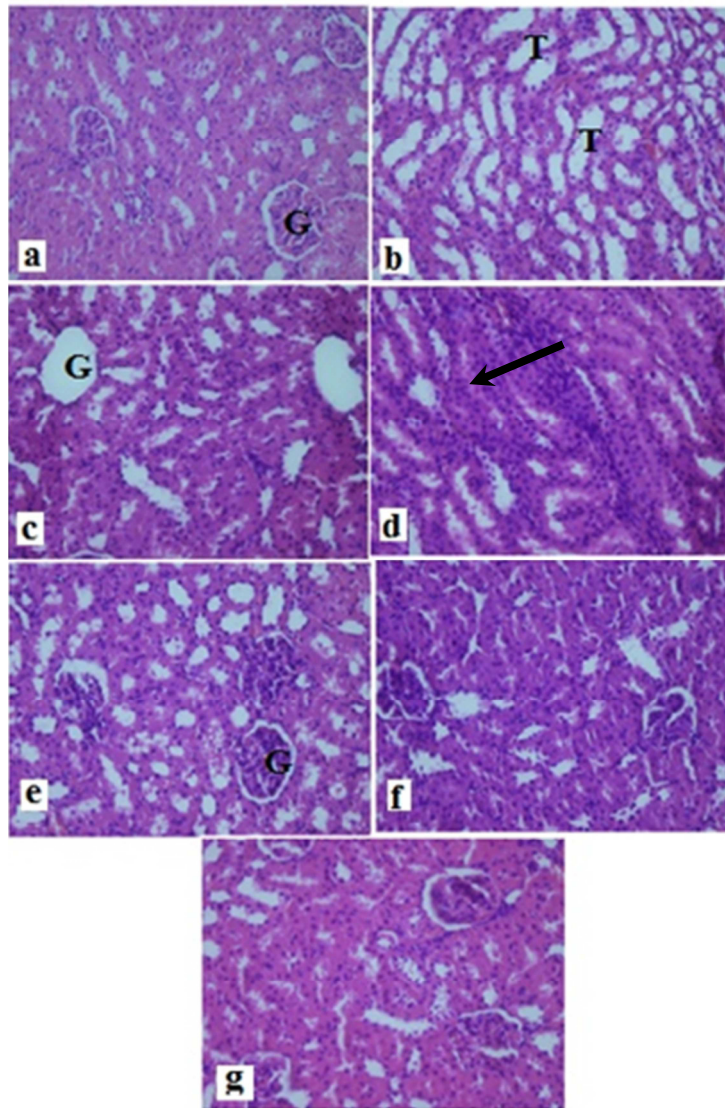
**Figure 3**  
Effect of daily oral administration of CFE and /or PDE on the level of renal angiogenic VEGF in diabetic rats.

**Table 3**  
Effect of CFE and /or PDE on renal function indices in serum of diabetic rat groups

Parameters	Control	Diabetes	Diabetes + CFE	Diabetes + PDE	Diabetes + CFE+ PDE
Uric acid	2.90 $\pm$ 0.18	9.6 $\pm$ 0.50 <sup>a</sup>	3.80 $\pm$ 0.12 <sup>c*S</sup>	3.52 $\pm$ 0.15 <sup>c*S</sup>	2.67 $\pm$ 0.23 <sup>*</sup>
Creatinine	0.59 $\pm$ 0.08	1.96 $\pm$ 0.153 <sup>a</sup>	0.89 $\pm$ 0.01 <sup>b*S</sup>	0.9 $\pm$ 0.04 <sup>b*S</sup>	0.68 $\pm$ 0.04 <sup>*</sup>
Urea	15.50 $\pm$ 2.02	48.9 $\pm$ 6.2 <sup>a</sup>	22.8 $\pm$ 0.90 <sup>c*</sup>	24.50 $\pm$ 2.5 <sup>c*</sup>	16.8 $\pm$ 0.93 <sup>*</sup>

Data are presented as mean  $\pm$ SD (n=10); <sup>a</sup>P  $\leq$  0.001, <sup>b</sup>P  $\leq$  0.01, <sup>c</sup>P  $\leq$  0.05 compared with the control group, <sup>\*</sup>P  $\leq$  0.001 compared with diabetic group, <sup>S</sup>P  $\leq$  0.05 compared with combination group (CFE+ PDE).





(a) Kidney picture of control rat showing normal renal glomeruli and tubules (b, c & d) Kidney pictures of diabetic rats, (b) showing severe vacuolar degradation of tubules; (c) showing glomerular degradation and (d) showing infiltration of inflammatory immune cells (arrow). (e, f & g) Kidney sections of diabetic rat treated with CFE, PDE C. and their combination (CFE+ PDE) respectively, showing more or less normal kidney architecture (H&E X 400).

**Figure 4**

***Kidney histomorphologic pictures of control and diabetic rats.***

## DISCUSSION

Extensive evidence revealed that DN is the most common reason of renal failing<sup>3</sup>. Drugs of plant origin is now accepted than the synthetic ones for controlling diabetic complexity for their safety and multifunctional roles. This investigation demonstrated the potential protective mechanism(s) of CFE and /or PDE versus renal damage induced by hyperglycemia in rats. To the extent of our information, this is the first work illustrating the protective impact of CFE and /or PDE versus renal damage in diabetic state. Significant elevation in the serum glucose level and a drop in insulin level were observed in STZC induced diabetic rats, indicating development of diabetic state in rats. Treatment with CFE and /or PDE markedly modulated the deviation in these parameters in

hyperglycemic rats with respect to diabetic untreated ones. The combination of the two plant extracts was the efficient in regulating these diabetic markers, suggesting that both plant extracts may improve the capability of diabetic rats to utilize the excess blood glucose by promoting the pancreatic beta-cells to produce more insulin. Similarly, the glycemic modulating efficacy of both *C. flexuosus* leaf extract and *P. dactylifera* fruit extract were previously reported<sup>12-13</sup>. In line with previous investigations, the current research illustrated that diabetic rats had decreased body weights and increased relative kidney weights compared with control ones, indicating renal hypertrophy<sup>4,19</sup>. The lowering in the body weights might be an indication of a decrease in the rate of protein synthesis coupled with an excessive breakdown of structural proteins as an alternative

source of energy due to the low availability of carbohydrates<sup>19</sup>. Renal hypertrophy may be due to the expansion of extracellular matrices which is prominent in diabetic nephropathy<sup>20</sup>. Administration of CFE and /or PDE appreciably restored the animal body weights and reduced the renal weight in the diabetic treated rat group. Treatment with the combination of CFE and PDE was the effective one in ameliorating the rat body weights and reducing the kidney hypertrophy. The pathogenicity of DN have been related to hyperglycemia induced chronic renal inflammation<sup>7</sup>. The event of renal inflammation can be initiated in DN by overexpression of proinflammatory molecules<sup>7,21</sup>. The present study showed up-regulation of serum CRP and renal TNF- $\alpha$  in diabetic rats with relation to non-diabetic ones. Elevation of such mediators in diabetic rats may be a mechanism by which diabetic hyperglycemia accelerates renal inflammatory deterioration. Some authors have reported that over-production of CRP during diabetes is one of the diabetic mechanism that lead to the progression of renal failure<sup>22</sup>. CRP induced by high glucose, significantly can up-regulate other inflammatory molecules, including TNF- $\alpha$ , and IL-1 $\beta$  via an NF- $\kappa$ B-dependent mechanism<sup>21</sup>. CRP can also promote the production of monocyte chemoattractant protein-1 (MCP-1) and fibrotic growth factors (such as TGF- $\beta$ 1, connective tissue growth factor) which synergistically able to cause renal inflammation and fibrosis<sup>22</sup>. On the other hand, it has reported that production of TNF- $\alpha$  is a toxic to renocytes and can cause their direct injury through inducing apoptosis and necrotic cell death<sup>23</sup>. Beside, reported toxic behavior of TNF- $\alpha$  on renocytes include the induction of transcription proteins, expression of cytokines and cell adhesion proteins, involved in the generation of other inflammatory molecules which collectively have an important action in renal failing in diabetes<sup>24</sup>. Our finding may propose that CRP and TNF  $\alpha$  act synergistically as inflammatory cofactors of high glucose to induce kidney failing. Thus, a prophylactic strategy that ameliorates the expression of inflammatory proteins could prevent organ dysfunction. Treatment of diabetic rats with CFE and /or PDE markedly reduced the concentration of TNF- $\alpha$  and CRP versus diabetic untreated animals. The possible mechanism by which the two plant extracts elicit anti-inflammatory effect in diabetic rats might be through inhibiting the production of inflammatory proteins. The suppressing effect of *P. dactylifera* fruit extract on the expression of inflammatory

cytokines has been demonstrated in an experimental animal model<sup>25</sup>. Also some dietary cucurbits show suppressing impact on the inflammatory proteins, namely interleukin (IL)-1 $\beta$  and TNF- $\alpha$  in sera of lipopolysaccharide (LPS)-inflamed mice<sup>26</sup>. Our work also demonstrated a pronounced increment in the angiogenic factor (VEGF) in the kidney of hyperglycemic animals. Similarly, excessive amount of VEGF was observed in response to hyperglycemia in diabetic rat<sup>27</sup>. VEGF is the primary angiogenic growth factor that promotes diabetic nephropathy in animals and human<sup>27-28</sup>. Overexpression of this factor can provoke inflammatory immune response and promote renal hypoxia, leading to proteinuria<sup>29</sup>. VEGF can cause thickening of tubular basement membrane, and renal interstitial fibrosis by stimulating extracellular matrix deposition, leading to impair in renal performance<sup>30-31</sup>. VEGF has been implicated in the suppression of endothelial NO production, which act as anticoagulant and anti-inflammatory factor, beside its role in vascular relaxation, thus causing vascular remodeling and inflammation<sup>32</sup>. The dramatic increase in renal VEGF in diabetic rats was significantly reduced by CFE and/or PDE treatment, indicating their beneficial anti-angiogenic effect. The combination of the two extracts was potential in reducing the angiogenic factor. To the best of found knowledge, this is the first work investigating the beneficial role of CFE against angiogenesis. However, previous study revealed the role of date fruit extract in reducing the expression of VEGF in experimentally induced liver damage<sup>25</sup>. Creatinine, uric acid and urea are the major biomarkers utilized of renal failing. The increment in these indices are potential pathological indicators of DN development<sup>33</sup>. Damaging in the renal tissue was evident with the elevation of these markers in serum of diabetic animals presented in this work. Histopathological examination of kidney picture of hyperglycemic animals showed obvious damages in the renal tissue as noticed by remarkable vacuolar degeneration of tubules, glomerular damages and infiltration of inflammatory immune cells. Treatment with the plant extracts, lonely or in a combination significantly abated the increased in kidney function biomarkers and reduced the aforementioned histo-cytological alterations, suggesting the renoprotective impact of the used extracts against diabetic nephropathy.

## CONCLUSION

Our work indicated that CFE and/or PDE exert protective impact versus DN by suppressing the induced inflammatory proteins and angiogenic factor, thus proposing that these plant extracts might have potential curative use for preventing and/or treatment of diabetes induced renal disorder.

## AUTHORS CONTRIBUTION STATEMENT

Amna A Saddiq planned the experiments, all authors carried out the experiment. Azza M Mohamed performed the statistical analysis of the results and drafted the article.

## REFERENCES

- Rathmann W, Giani G. Global Prevalence of Diabetes: Estimates for the Year 2000 and Projections for 2030: Response to Wild et al. *Diabetes Care*. 2004;27(10):2568–9. DOI:10.2337/diacare.27.10.2568
- Kitada M, Zhang Z, Mima A, King GL. Molecular mechanisms of diabetic vascular complications. *J Diabetes Investig*. 2010;1(3):77–89. DOI:10.1111/j.2040-1124.2010.00018.x
- Gheith O, Farouk N, Nampoory N, Halim MA A-OT. Diabetic kidney disease: World Wide difference of prevalence and risk factors. *J Nephroarmacol*. 2015;5(1):49–56. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5297507/>
- Olatunji OJ, Chen H, Zhou Y. Lycium chinense leaves extract ameliorates diabetic nephropathy by suppressing hyperglycemia mediated renal oxidative stress and inflammation. *Biomed Pharmacother*. 2018;102:1145–51. DOI:10.1016/j.biopha.2018.03.037
- Skupien J, Warram JH, Smiles AM, Stanton RC, Krolewski AS. Patterns of Estimated Glomerular Filtration Rate Decline Leading to End-Stage Renal Disease in Type 1 Diabetes. *Diabetes Care*. 2016;39(12):2262–9. DOI:10.2337/dc16-0950
- Brownlee M. The Pathobiology of Diabetic Complications: A Unifying Mechanism. *Diabetes*. 2005;54(6):1615–25. DOI:10.2337/diabetes.54.6.1615
- Wan T-T, Li X-F, Sun Y-M, Li Y-B, Su Y. Recent advances in understanding the biochemical and molecular mechanism of diabetic retinopathy. *Biomed Pharmacother*. 2015;74:145–7. DOI:10.1016/j.biopha.2015.08.002
- Dhiman K, Gupta A, Sharma DK, Gill NS, Goyal A. A Review on the Medicinally Important Plants of the Family Cucurbitaceae. *Asian J Clin Nutr*. 2012;4(1):16–26. DOI:10.3923/ajcn.2012.16.26
- Ma C, Yu H, Xiao Y, Wang H. Momordica charantia extracts ameliorate insulin resistance by regulating the expression of SOCS-3 and JNK in type 2 diabetes mellitus rats. *Pharm Biol*. 2017; 55(1):2170-7. DOI:10.1080/13880209.2017.1396350
- Joseph B, Jini D. Antidiabetic effects of Momordica charantia (bitter melon) and its medicinal potency. *Asian Pacific J Trop Dis*. 2013;3(2):93–102. DOI:10.1016/s2222-1808(13)60052-3
- Mendi YY, Eldoğan S, Gutakev R, İpek M, Çürük P CS. Regeneration and histological analysis of snake melon (*Cucumis melo* var. *flexuosus* L. Naudin) by direct organogenesis. *Turk J Agric*. 2010; 34(4):309–14. Available from: <http://journals.tubitak.gov.tr/agriculture/issue/s/tar-10-34-4/tar-34-4-5-0905-8.pdf>
- Ibrahim DS. Neuroprotective effect of *Cucumis melo* Var. *flexuosus* leaf extract on the brains of rats with streptozotocin-induced

## ACKNOWLEDGMENT

The authors gratefully acknowledge the Scientific Chair of YA Jameel of the Prophetic Medical Application, Jeddah, Kingdom of Saudi Arabia, for the financial support of this work in the form of a research project.

## FUNDING ACKNOWLEDGEMENT

We acknowledge the resources and financial support for the study provided by the Scientific Chair of YA Jameel of the Prophetic Medical Application, Jeddah, Kingdom of Saudi Arabia.

## CONFLICT OF INTEREST

Conflict of interest declared none.

- diabetes. *Metab Brain Dis.* 2016;32(1):69–75. DOI:10.1007/s11011-016-9886-y
13. Michael HN, Salib JY, Eskander EF. Bioactivity of Diosmetin Glycosides Isolated from the Epicarp of Date Fruits, *Phoenix dactylifera*, on the Biochemical Profile of Alloxan Diabetic Male Rats. *Phyther Res.* 2012;27(5):699–704. DOI:10.1002/ptr.4777
  14. Rahmani AH, Aly SM, Ali H, Babiker AY, Srikar S KA. Therapeutic effects of date fruits (*Phoenix dactylifera*) in the prevention of diseases via modulation of anti-inflammatory, anti-oxidant and anti-tumour activity. *Int J Clin Exp Med.* 2014;7(3):483–91. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3992385/>
  15. Zhang C-R, Aldosari SA, Vidyasagar PSP V, Nair KM, Nair MG. Antioxidant and Anti-inflammatory Assays Confirm Bioactive Compounds in Ajwa Date Fruit. *J Agric Food Chem.* 2013;61(24):5834–40. DOI:10.1021/jf401371v
  16. Mallhi TH, Qadir MI, Ali M, Ahmad B, Khan YH RA. Ajwa date (*Phoenix dactylifera*): an emerging plant in pharmacological research. *Pak J Pharm Sci.* 2014;27(3):607–16.
  17. Wu KK, Huan Y. Streptozotocin-Induced Diabetic Models in Mice and Rats. In: *Current Protocols in Pharmacology.* John Wiley & Sons, Inc.; 2008. DOI:10.1002/0471141755.ph0547s40
  18. Mard SA, Jalalvand K, Jafarinejad M, Balochi H NM. Evaluation of the antidiabetic and antilipaemic activities of the hydroalcoholic extract of *Phoenix Dactylifera* palm leaves and its fractions in alloxan-induced diabetic rats. *Malaysian J Med Sci.* 2010;17(4):4–13. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3216186/>
  19. Murray AJ, Anderson RE, Watson GC, Radda GK, Clarke K. Uncoupling proteins in human heart. *Lancet.* 2004;364(9447):1786–8. DOI:10.1016/s0140-6736(04)17402-3
  20. Zhang C, Li Q, Lai S, Yang L, Shi G, Wang Q, et al. Attenuation of diabetic nephropathy by Sanziguben Granule inhibiting EMT through Nrf2-mediated anti-oxidative effects in streptozotocin (STZ)-induced diabetic rats. *J Ethnopharmacol.* 2017;205:207–16. DOI:10.1016/j.jep.2017.05.009
  21. Mezzano S, Aros C, Droguett A, Burgos ME, Ardiles L, Flores C, et al. NF- $\kappa$ B activation and overexpression of regulated genes in human diabetic nephropathy. *Nephrol Dial Transplant.* 2004;19(10):2505–12. DOI:10.1093/ndt/gfh207
  22. Liu F, Chen HY, Huang XR, Chung ACK, Zhou L, Fu P, et al. C-reactive protein promotes diabetic kidney disease in a mouse model of type 1 diabetes. *Diabetologia.* 2011;54(10):2713–23. DOI:10.1007/s00125-011-2237-y
  23. Laster SM, Wood J G GL. Tumor necrosis factor can induce both apoptotic and necrotic forms of cell lysis. *J Immunol.* 1988;141(8):2629–34. Available from: <http://www.jimmunol.org/content/141/8/2629>
  24. Ortiz A, Bustos C, Alonso J, Alcázar R, López-Armada MJ, Plaza JJ, González E EJ. Involvement of tumor necrosis factor-alpha in the pathogenesis of experimental, and human glomerulonephritis. *Adv Nephrol Necker Hosp.* 1995;24:53–77. Available from: <https://europepmc.org/abstract/med/7572422>
  25. Al-Rasheed NM, Attia HA, Mohamad RA, Al-Rasheed NM, Al-Amin MA, AL-Onazi A. Aqueous Date Flesh or Pits Extract Attenuates Liver Fibrosis via Suppression of Hepatic Stellate Cell Activation and Reduction of Inflammatory Cytokines, Transforming Growth Factor- $\beta$ 1 and Angiogenic Markers in Carbon Tetrachloride-Intoxicated Rats. *Evidence-Based Complement Altern Med.* 2015;2015:1–19. DOI:10.1155/2015/247357
  26. Hamad I, AbdElgawad H, Al Jaouni S, Zinta G, Asard H, Hassan S, et al. Metabolic Analysis of Various Date Palm Fruit (*Phoenix dactylifera* L.) Cultivars from Saudi Arabia to Assess Their Nutritional Quality. *Molecules.* 2015;20(8):13620–41. DOI:10.3390/molecules200813620
  27. Cooper ME, Vranes D, Youssef S, Stacker SA, Cox AJ, Rizkalla B, et al. Increased renal expression of vascular endothelial growth factor (VEGF) and its receptor VEGFR-2 in experimental diabetes. *Diabetes.* 1999;48(11):2229–39. DOI:10.2337/diabetes.48.11.2229
  28. Kanasaki Y, Suzuki D, Uehara G, Toyoda M, Katoh T, Sakai H, et al. Vascular endothelial growth factor gene expression is correlated with glomerular neovascularization in human diabetic

- nephropathy. *Am J Kidney Dis.* 2005; 45(2):288–94.  
DOI:10.1053/j.ajkd.2004.09.020
29. Lin S, Teng J, Li J, Sun F, Yuan D, Chang J. Association of Chemerin and Vascular Endothelial Growth Factor (VEGF) with Diabetic Nephropathy. *Med Sci Monit.* 2016;22:3209–14.  
DOI:10.12659/msm.896781
30. Cheng H, Harris R. Renal Endothelial Dysfunction in Diabetic Nephropathy. *Cardiovasc Hematol Disord Targets.* 2014;14(1):22–33.  
DOI:10.2174/1871529x14666140401110841
31. Ma L, Gao Y, Chen G, Gong J, Yang D, Xie Y, Wang M, Chen H SM. Relationships of urinary VEGF/CR and IL-6/CR with glomerular pathological injury in asymptomatic hematuria patients. *Med Sci Monit.* 2015;21:356–62. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4321409/>
32. Nakagawa T, Sato W, Glushakova O, Heinig M, Clarke T, Campbell-Thompson M, et al. Diabetic Endothelial Nitric Oxide Synthase Knockout Mice Develop Advanced Diabetic Nephropathy. *J Am Soc Nephrol.* 2007;18(2):539–50.  
DOI:10.1681/asn.2006050459
33. Mestry SN, Dhodi JB, Kumbhar SB, Juvekar AR. Attenuation of diabetic nephropathy in streptozotocin-induced diabetic rats by *Punica granatum* Linn. leaves extract. *J Tradit Complement Med.* 2017;7(3):273–80.  
DOI:10.1016/j.jtcme.2016.06.008